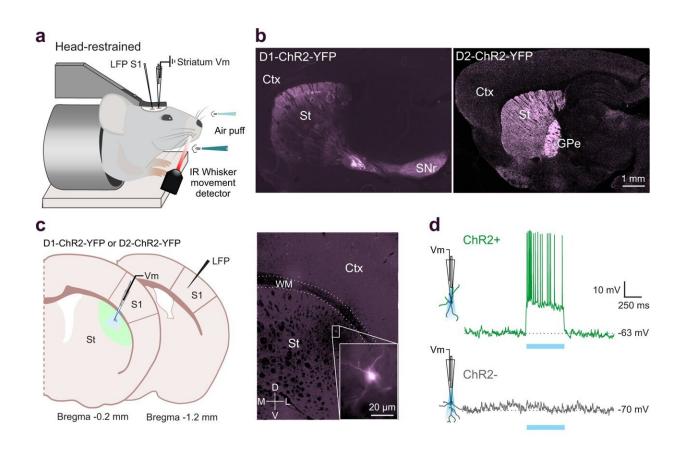


Movement shown to reduce sensory responses in Parkinson's disease

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Whole-cell recordings from opto-tagged MSNs in the DLS of awake behaving mice. a Schematic of the experimental setup. Whole-cell recordings were performed in the mouse DLS with the optopatcher for online optogenetic classification, simultaneously with LFP recordings in S1. Whiskers were stimulated by air-puffs delivered independently to ipsilateral and contralateral whiskers, and whisking behavior was monitored using a non-contact infrared LED-photodiode. b Opto-tagging of MSNs was obtained using either D1-ChR2-YFP (labeling dMSNs, left) or D2-ChR2-YFP (labeling iMSNs, right)



mice (n = 15 mice for D1, n = 19 mice for D2). Typical projections from dMSNs in D1-ChR2-YFP mice to SNr and from iMSNs to the GPe in the D2-ChR2-YFP mice are evident. Scale bar, 1 mm, c Schematic representation showing whole-cell and LFP recording locations from Bregma (left). The image and magnified inset (right) show an example (from 34 independent neurons with similar results) of a biocytin-filled MSN from DLS following an in vivo wholecell recording in a behaving mouse. The inset shows the same cell in higher magnification. d Opto-tagging of MSNs using the optopatcher. Depolarizing responses to photostimulation of a positive cell (ChR2 +) in D1-ChR2-YFP mouse (top) for the duration of the stimulation. Negative cells (ChR2 – , bottom) did not respond with depolarization to photostimulation. e Example of spontaneous membrane potential activity in an opto-tagged iMSN, whole-cell recorded in the DLS of an awake behaving mouse. Membrane potential (Vm, red) of the neuron was recorded simultaneously with measurement of whisker activity (Whisker, green), air puff stimulation indicated in gray (ipsi/contra). f Synaptic responses of the neuron showed in e, to contra- and ipsilateral whisker deflection, g Depolarization of the membrane potential preceded whisker movement in the same neuron showed in e. h Venn diagram showing the number of MSNs responding to whisker stimulation (turquoise circle) and to spontaneous whisking movement (pink circle) or absence of both (red circle). LFP Local Field Potential, Barrel field somatosensory cortex S1, IR infrared light, SNr Substantia Nigra pars reticulata, GPe Globus Pallidus externa, Ctx Cortex, WM White Matter, St Striatum, ChR2 Channelrhodopsin, a.u. arbitrary units. Credit: Nature Communications (2023). DOI: 10.1038/s41467-023-36648-0

Researchers at Karolinska Institutet have been looking at how movement shapes our senses and how sensory and motor processes are both affected in Parkinson's disease. In a study published in *Nature Communications*, de la Torre-Martinez et al, show that movement reduces the responses to sensory input and that both sensory and motor processes are altered in an animal model of Parkinson's disease.

Movement control and sensory integration are the fundamental functions



of our nervous system. Within the <u>brain</u>, a group of deep nuclei called the basal ganglia are involved in such sensorimotor functions. The basal ganglia are also the primary brain region affected in Parkinson's disease.

"The basal ganglia are involved in both sensory and motor functions and we show here how <u>movement</u> shapes sensory responses," says Gilad Silberberg, professor at the Department of Neuroscience at Karolinska Institutet and corresponding author of the article. "We also show that in Parkinsonian mice both sensory integration and movement-related responses are changed."

Most previous work in the basal ganglia field focused on motor impairments, while here, the researchers highlight the interactions between motor and sensory functions.

"The results are important for our understanding of sensory-motor interactions in normal brain function. We show that single neurons in the basal ganglia respond to both sensory stimulation and movement. We also show that movement reduces the strength of sensory inputs and we describe the mechanisms that make it happen," says Roberto de la Torre-Martinez, postdoctoral researcher and co-author of the study.

Electrical recording of deep brain neurons

The researchers used a special electrical recording method called patchclamp electrophysiology, which received the Nobel Prize in 1991.

"Here, we used this method to record deep brain neurons in awake mice while they integrated <u>sensory inputs</u>. These recordings are technically challenging but they can provide information about the synaptic inputs to neurons while the animal is engaged in sensory-motor functions. We used healthy mice and compared their sensory-motor properties with Parkinsonian (dopamine-depleted) mice," explains Maya Ketzef,



assistant professor at the same department and co-author of the study.

The next step is to combine the patch-clamp recordings with imaging of different neuromodulators in the basal ganglia.

"Traditionally, dopamine has been the main neuromodulator studied in the <u>basal ganglia</u>, and we would like to study how dopamine interacts with other neuromodulators such as acetylcholine and serotonin," says Gilad Silberberg.

More information: Roberto de la Torre-Martinez et al, Ongoing movement controls sensory integration in the dorsolateral striatum, *Nature Communications* (2023). DOI: 10.1038/s41467-023-36648-0

Provided by Karolinska Institutet

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